



# Analyzing structural variation in breast cancer with long read sequencing of patient-derived organoids

#### Rachel M Sherman

Johns Hopkins University Schatz and Salzberg Labs Biological Data Science 2018

#### Structural variation characterizes cancer

#### Prognostic indicator

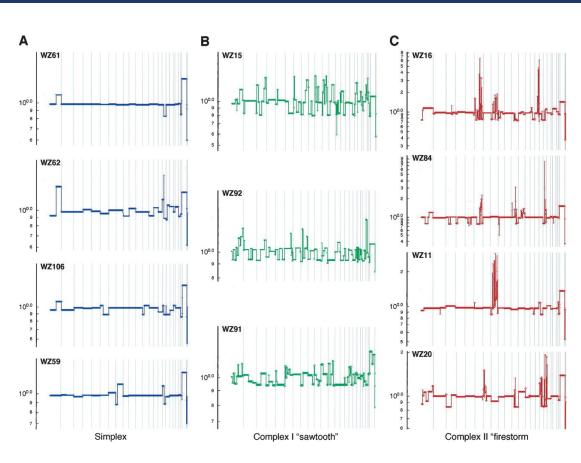
Greater instability leads to worse patient outcomes

#### Copy number changes

Amplifications and deletions, especially of oncogenes and tumor suppressors

#### Gene fusions

Can drastically alter gene expression levels



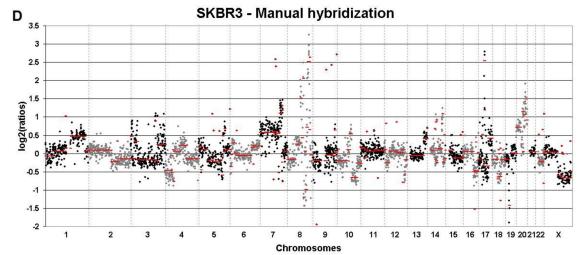
**Figure 2.** Major types of tumor genomic profiles. Segmentation profiles for individual tumors representing each category: (*A*) simplex; (*B*) complex type I or sawtooth; (*C*) complex type II or firestorm. Scored events consist of a minimum of six consecutive probes in the same state. The *y*-axis displays the geometric mean value of two experiments on a log scale. Note that the scale of the amplifications in *C* is compressed relative to *A* and *B* owing to the high levels of amplification in firestorms. Chromosomes 1–22 plus X and Y are displayed in order from *left* to *right* according to probe position.

(Hicks et al, 2006, Genome Research)

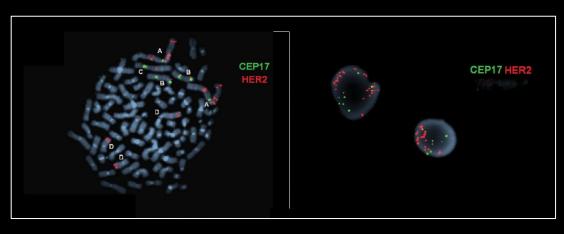
#### SK-BR-3 Her2 amplified cell line

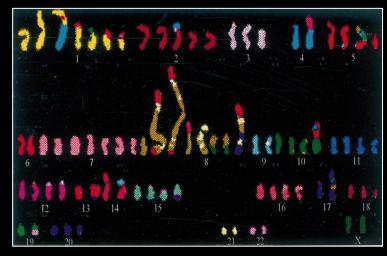
Derived in 1970, most commonly used Her2 amplified cell line

FISH, aCGH, and karyotyping show large-scale changes



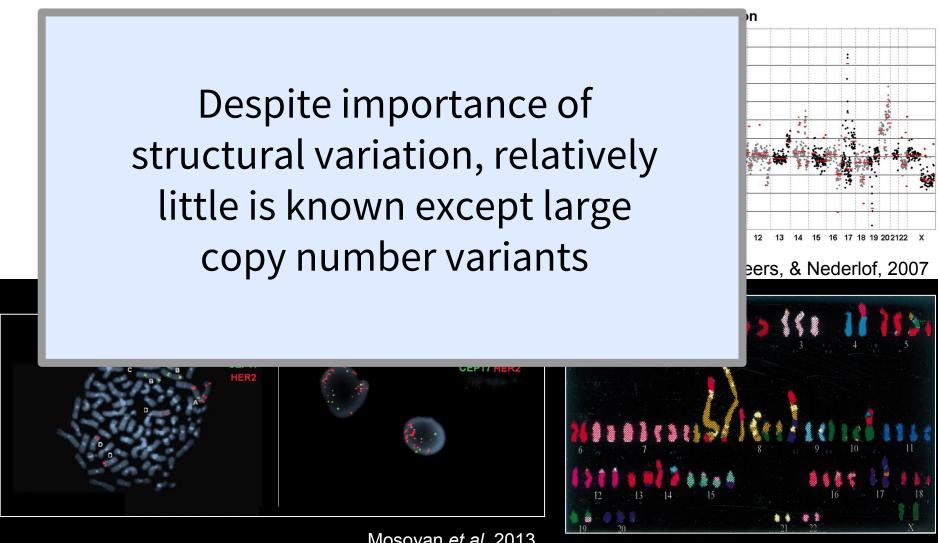
Joosse, Beers, & Nederlof, 2007





Mosoyan et al, 2013

## SK-BR-3 Her2 amplified cell line

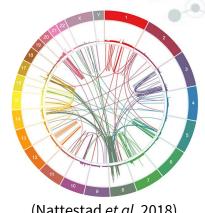


# Analyzing variation with long reads

#### **Part 1 (SK-BR-3)**

How prevalent are mid-sized variants?

How necessary are long reads?



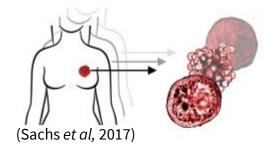
(Nattestad et al, 2018)

#### Part 2 (Patient-derived organoids)

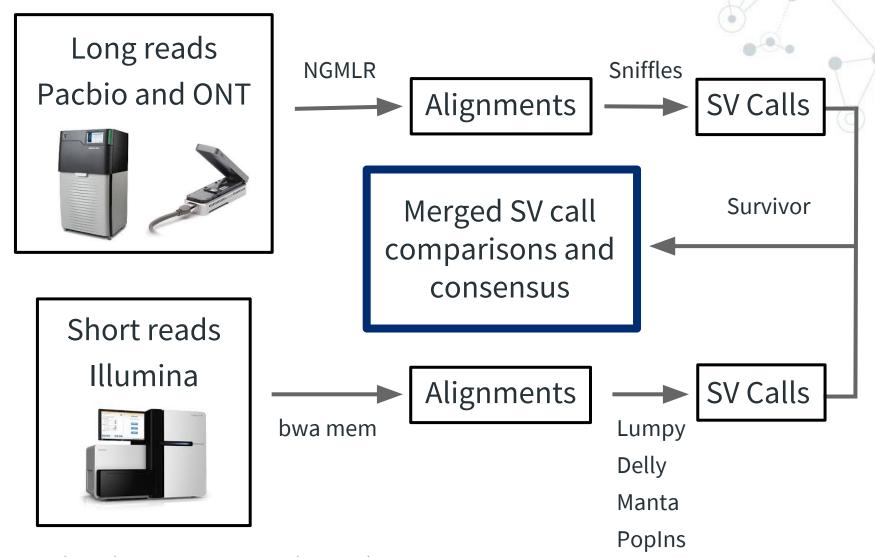
What new variants have we found?

How do they compare to a cell line (SK-BR-3)?

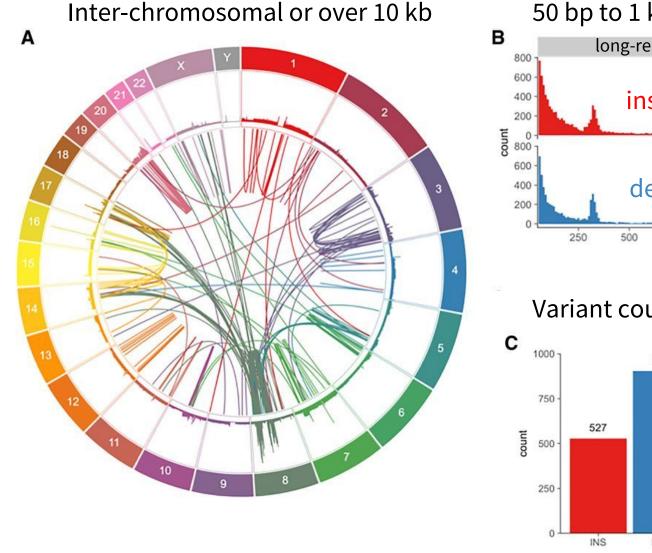
What else can long reads tell us?



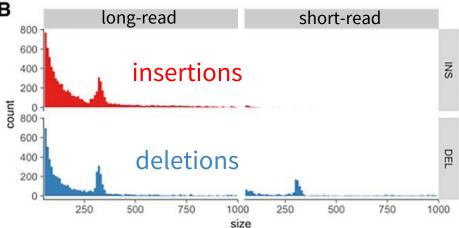
# SK-BR-3 sequencing and analysis



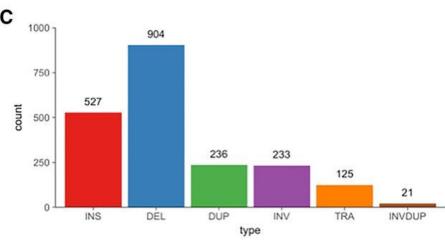
(Nattestad *et al*, Genome Research, 2018)



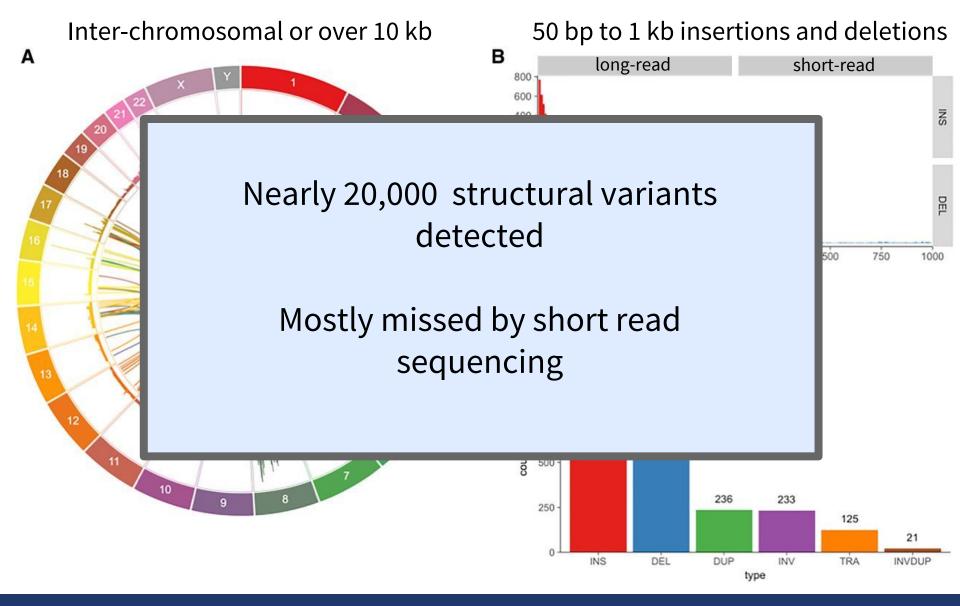
#### 50 bp to 1 kb insertions and deletions



#### Variant counts over 1 kb

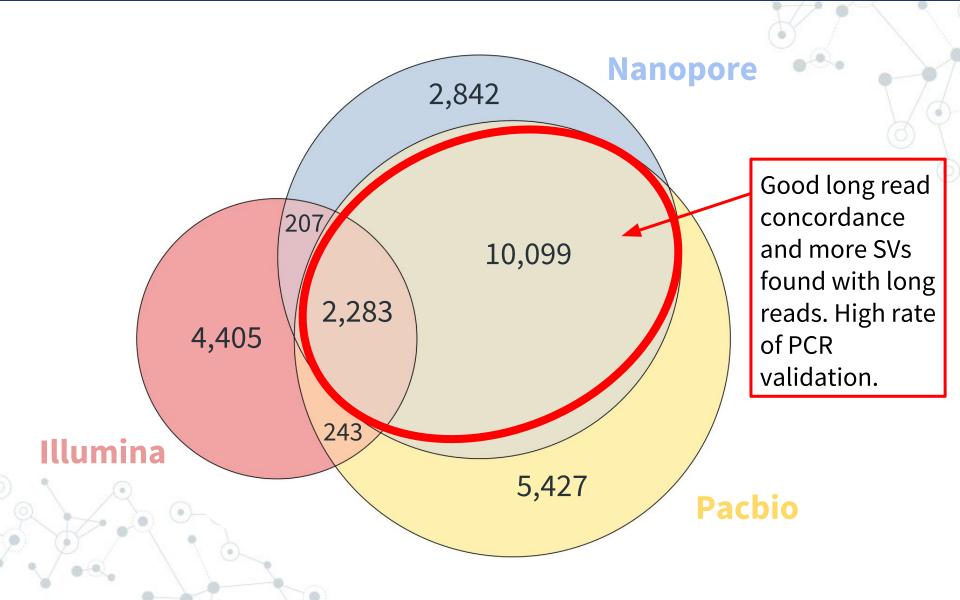


Structural variants detected with PacBio in SKBR3 (Nattestad et al, 2018).

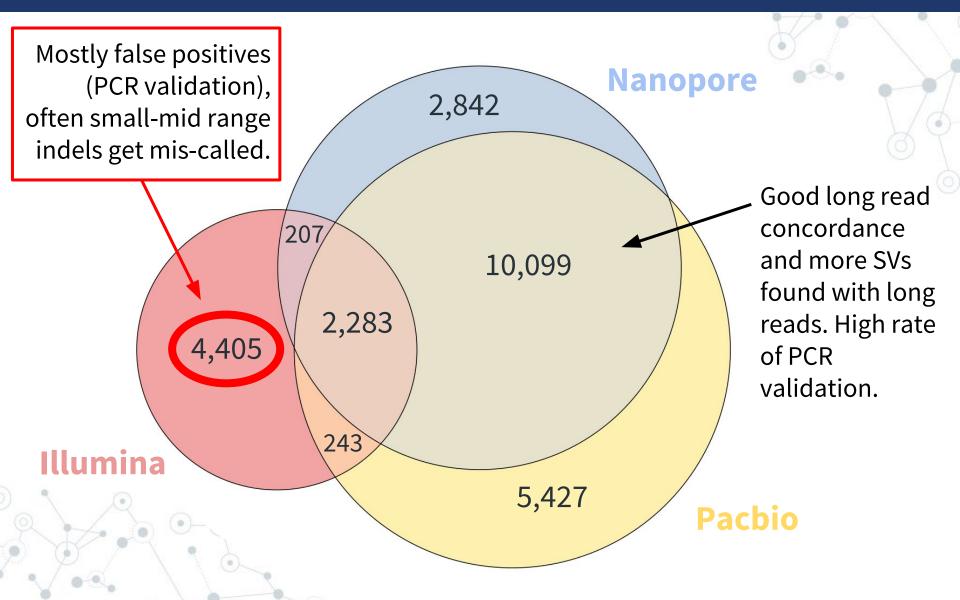


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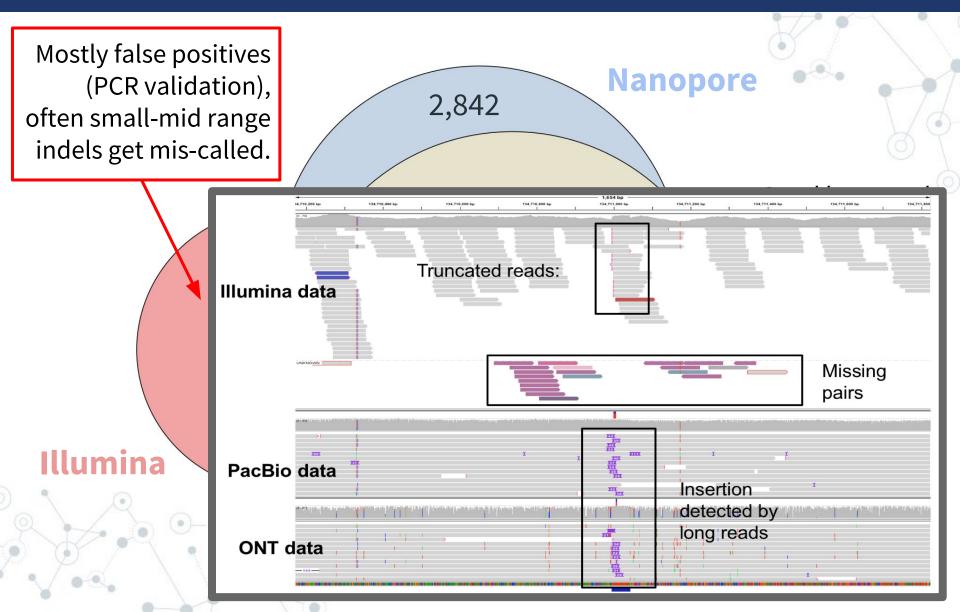
# Variant calling across platforms



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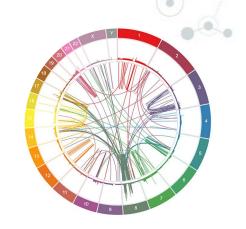


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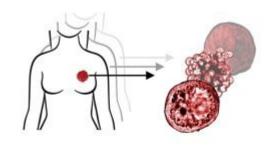


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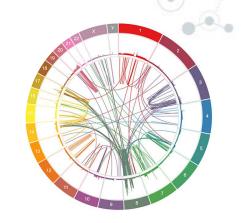
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# Analyzing variation with long reads

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#### Part 2 (Patient-derived organoids)

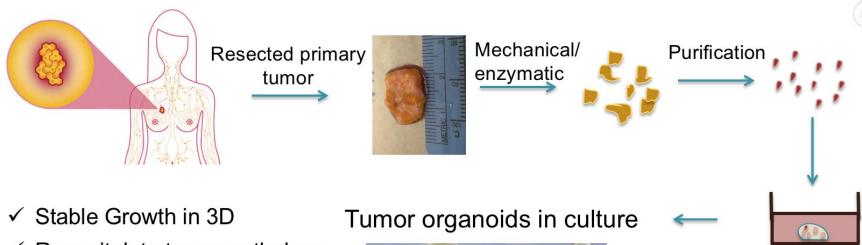
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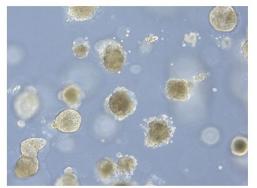
What else can long reads tell us?

# Patient-derived tumor organoids

Breast cancer patient tumor organoid, ER+, PR+, Her2-



- ✓ Recapitulate tumor pathology & treatment response
- Maintenance of tissue/tumor heterogeneity
- ✓ "2017 Method of the Year" Nature Methods



Plating on Matrigel
Add growth factors

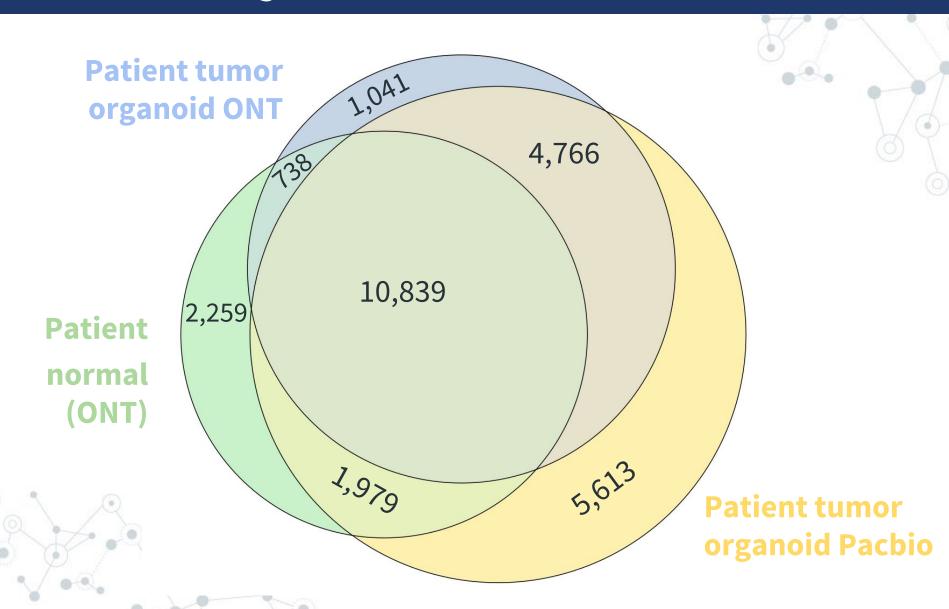


# Samples and sequencing types

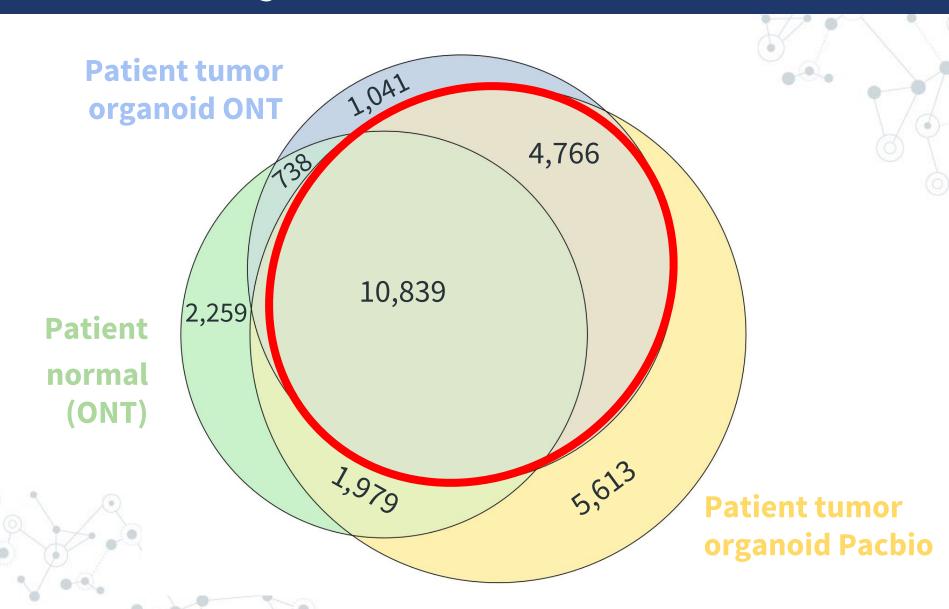
	ONT	Pacbio	Illumina
Patient tumor (organoid, ER+, PR+, Her2-)			
SK-BR-3 (cell line, ER-, PR-, Her2+)	/	/	
MCF-7 (cell line, ER+, PR+, Her2-)			
MDA-MB-231 (cell line, ER-, PR-, Her2-)			

# Patient normal (breast tissue) MCF-10A (normal breast cell line) NA12878 HG002 (Genome in a Bottle)

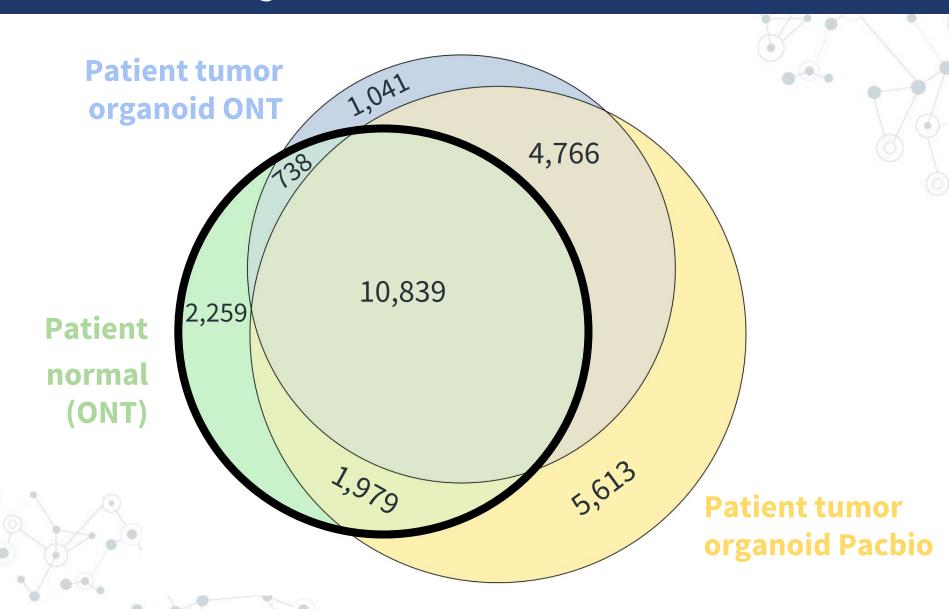
# Tumor organoid vs normal variant calls



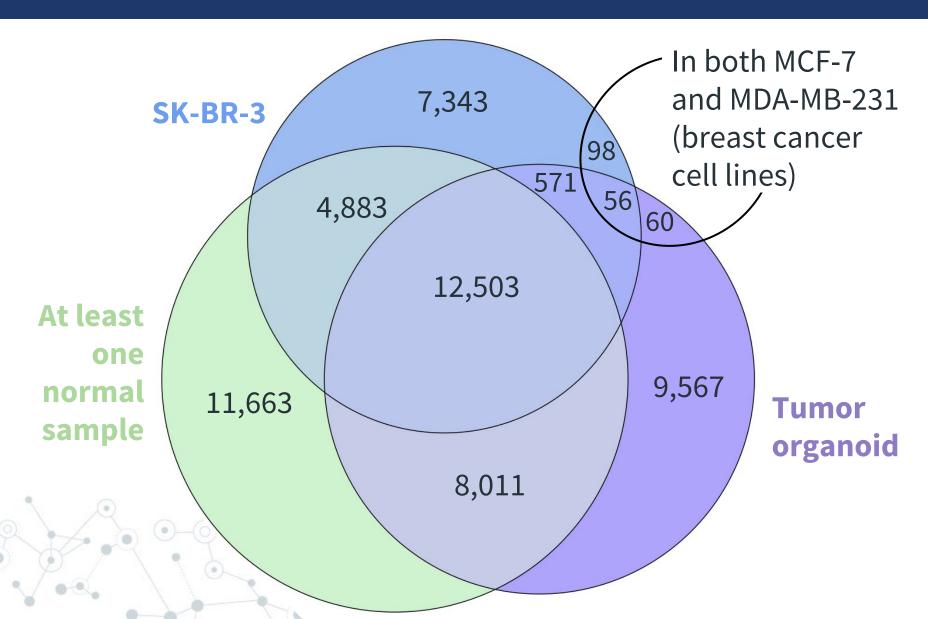
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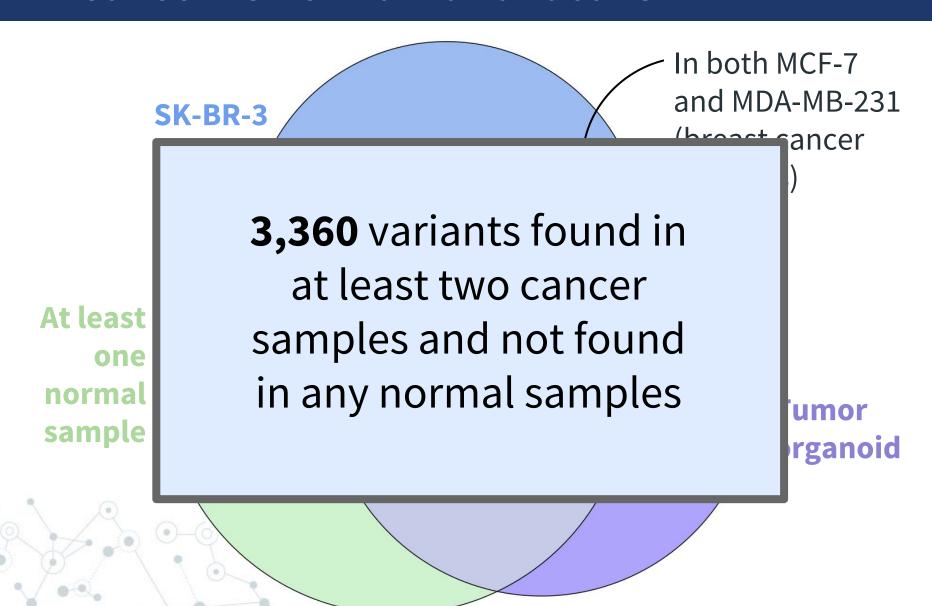
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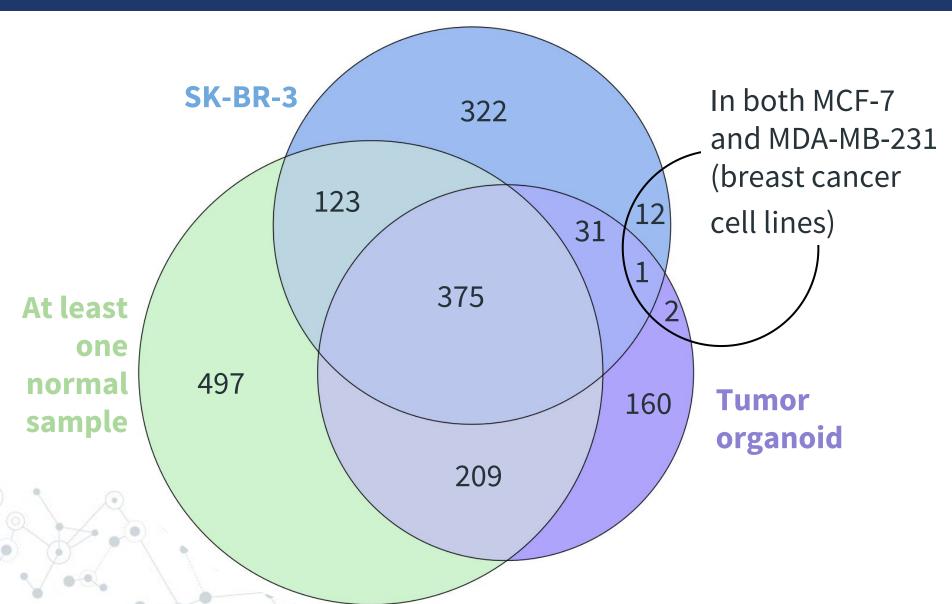
#### Cancer vs normal variant calls



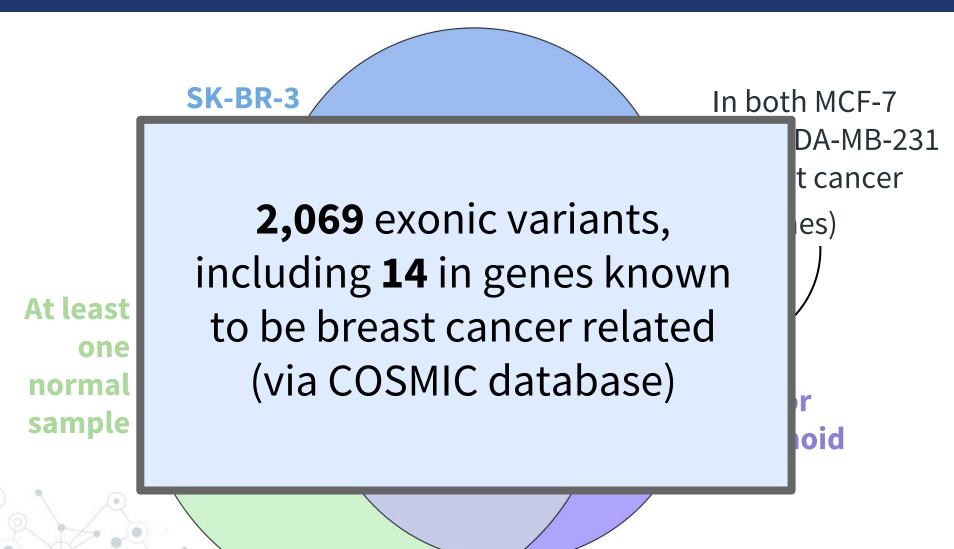
#### Cancer vs normal variant calls



# Exonic variants only

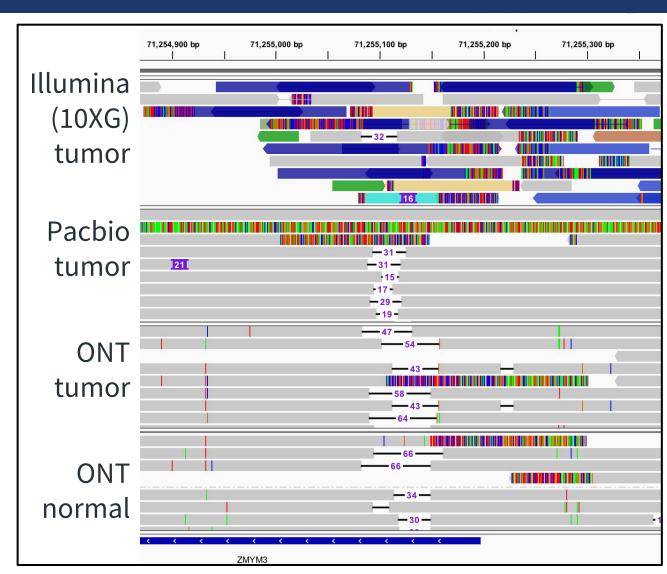


# Exonic variants only



#### ~50 bp deletion in ZMYM3 exon

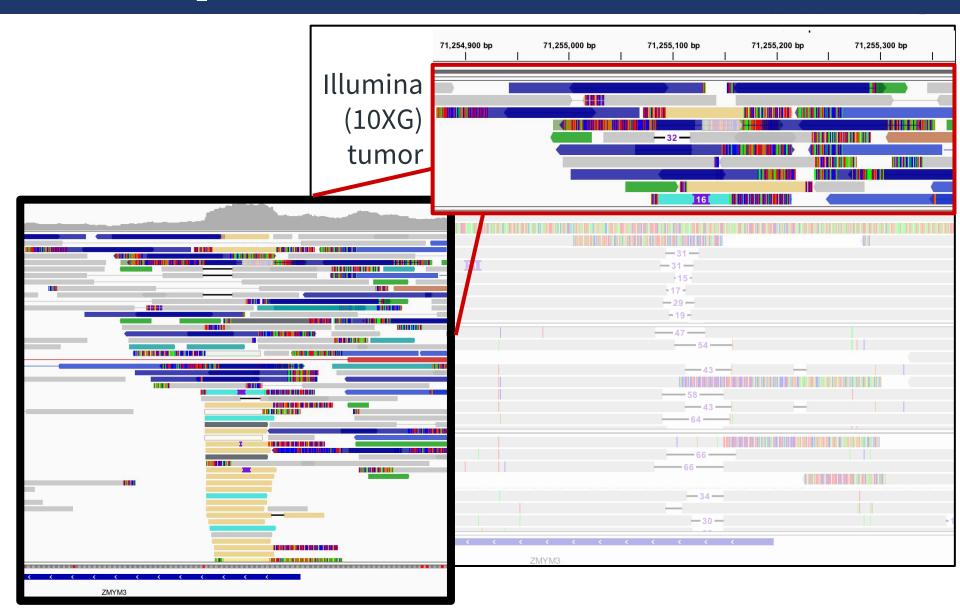
- Evidence it
   promotes DNA
   repair via BRCA1
   recruitment <sup>1</sup>
- © Encodes part of a histone deacetylasecontaining complex involved in gene silencing\*



<sup>\*</sup> via Genecards 

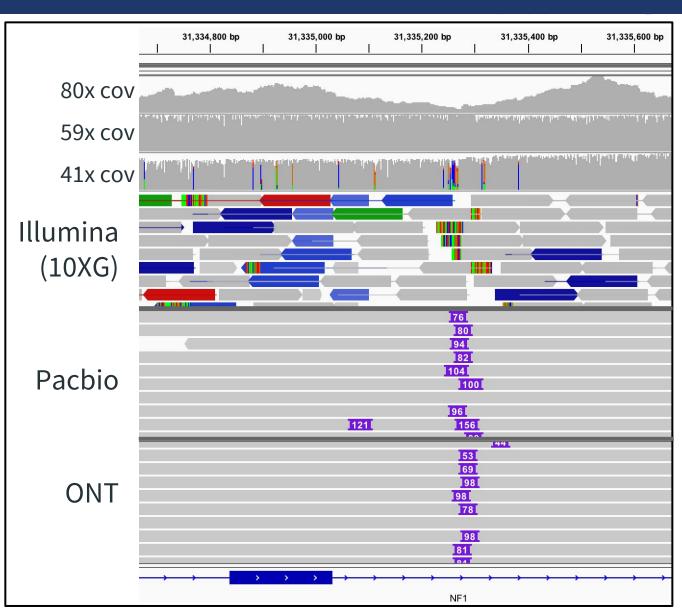
<sup>1</sup> Leung, J. W., et al (2017).

# ~50 bp deletion in ZMYM3 exon



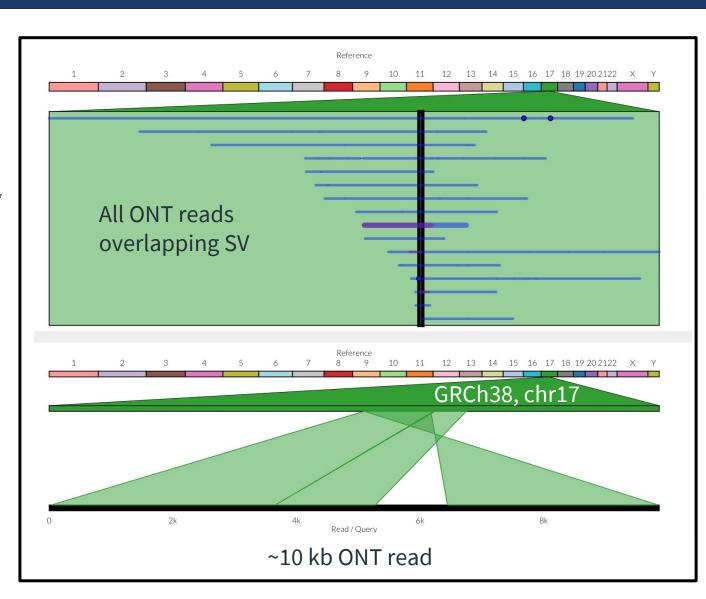
#### ~100 bp insertion in NF1 intron

- Negative
   regulator of the
   ras signal
   transduction
   pathway \*
- Mown to be involved in the "Integrated Breast Cancer Pathway"



# Inverted duplication in ERBB2 (Her2) exon

- Amplification/ overexpression detected in 20-30% of breast cancers\*
- Involved in transcriptional regulation \*



#### Conclusions and future directions

# Long reads are critical to variant discovery!

Long term: Work towards collecting long read data sets

Immediate: Get more out of short read data, given a small collection of long read samples

Many short read samples

Paragraph genotyper <sup>1</sup>

Few long read samples SV calling Long read SV catalog

SVs found in short read samples

<sup>&</sup>lt;sup>1</sup> https://github.com/Illumina/paragraph

# Acknowledgments

#### **Johns Hopkins University**

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Melissa Kramer

Robert Wappel

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#### Illumina (Paragraph)

Peter Krusche

Sai Chen

Michael Eberle

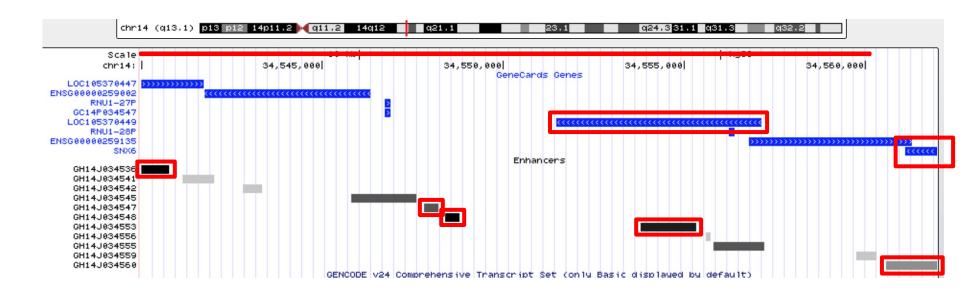
Felix Schlesinger

Egor Dolzhenko



# Questions?

# Exonic variant (all cancer, no normal samples)

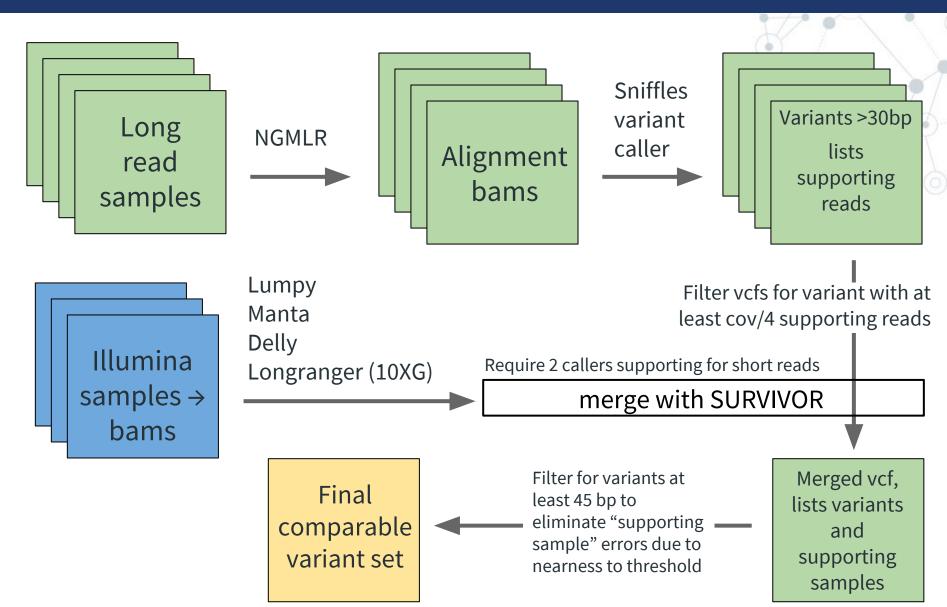


17 kb inversion

Encompases ncRNA LOC105370449

Just ahead of SNX6, aka TRAF4, a known oncogene

#### Protocol for variant comparison



# Data statistics for all samples

Sample	Sequencing technology	Tumor or normal	Average coverage	Number of supporting reads required for Sniffles calls
SK-BR-3	ONT minion		27.8	7
	ONT promethion	Tumor	25	6
	Pacbio	Turrior	52.9	10
	Illumina		20	N/A
MCF7	ONT minion	Tumor	16.5	4
MDA-MB-231	ONT minion	Tumor	17	4
Patient Tumor Organoid	10X Genomics		28	N/A
	ONT minion	Tumor	31.5	8
	Pacbio		55.3	10
Patient Breast Tissue	ONT minion	Normal	11.6	3
MCF10A	ONT minion	Normal	22.4	6
HG002 (GiaB son)	Pacbio	Normal	50.4	10
NA/GM12878	ONT minion		31.2	8
	Pacbio	Normal	41.4	10
	ONT minion		20	5

# Platform concordance in tumor organoid

